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Endocrinal toxicity of industrial solvents – A mini review

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Endocrine system can be affected by various organic compounds. The review describes the effects of major industrial solvents on adrenal, thyroid and parathyroid glands in man and experimental animals. Further, their toxicity in pancreas, pituitary, testis and ovary has also been discussed. An attempt has been made to offer a historical and general information on solvent toxicity in endocrine glands. Possible mechanisms, in brief, have also been discussed. Endocrine toxicity caused by industrial solvents deserves more attention than hitherto paid. An understanding of hormonal disorders caused by industrial solvents will be important from occupational health point of view.

Keywords: Adrenal, Endocrinal disruption, Gonads, Industrial solvents, Thyroid

Outline

Over the last two decades, there has been increasing scientific concern and public debate regarding the adverse effects of chemical pollutants present in the environment that can interfere with the normal functioning of the endocrine system in wildlife and in humans (the so-called endocrinedisrupting chemicals, EDCs). These concerns have been fuelled primarily by reports of disrupted reproductive function and development in certain wild-mammals, birds, fish, amphibians, mollusks and by the increased incidence of certain diseases of the endocrine system in humans. Some of the adverse effects observed in several species are strongly associated with exposure to chemicals that mimic or interfere with hormone function, particularly estrogen function, but in many cases, the causal link between exposure to EDCs and endocrine disruption is unclear. Because of the diverse effects of EDCs on the thyroid, retinoid. androgen, estrogen, and corticosteroid systems of a wide range of animals, it is imperative to address the extent of the risk posed by EDCs to wildlife and man. The ecological relevance of endocrine disruption is, however, difficult to quantify, as there is limited understanding of how physiological changes affect the individual animal

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and how individual responses affect population and community. Further, a major challenge faced by environmental biologists is the need to place endocrine disruption into context with other environmental pressures faced by populations, viz. global warming and climate change.

Introduction

An endocrine gland is composed of a prominent mass or parenchyma of secretary cells as well as connective tissue, blood vessels and nerves. Endocrine glands secrete their products (hormones) directly into the bloodstream. Hormones were originally considered to be synthesized within specific endocrine organs and then secreted into the bloodstream to act on specific target tissues distance away to evoke a specific metabolic/physiological response¹. Hormones activate very specific cells or target tissues. The identification of hormones in single cell organisms confirms the specificity of hormones to particular biochemical pathways within a cell. This specificity of hormones is achieved by finally tuned regulation of specific receptors for each hormone. Receptors response is highly specific. Receptors are found in membranes and within the cytoplasm of cells. There are two principal pathways for receptor activation. Firstly, there are receptors on the plasma membrane for protein and peptide hormones. Secondly, there are cytoplasmic or nuclear receptors for lipophilic moieties such as thyroid and steroid hormones. Many receptors for endocrine target organs

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are ligand-dependent transcriptional regulators that belong to a super family of proteins that include oestrogens. androgens, retinoic acid, thyroid hormones and vitamin D. These hormones, having molecular weights between 300 and 400 Da, can permeate the plasma membrane. Once they enter the cell, these hormones can bind to specific protein both in the cytoplasm or the nucleus². The mechanism of action of hormones has been categorized into two groups. The hormones in group 1 are lipophilic. These hormones bind to intracellular receptors in the cytoplasm or the nucleus and cause confirmational changes. The hormone-receptor complex then binds to a specific region on DNA called the hormone response element (HRE). This interaction with the help of various accessory factors and co-regulators, results in the activation or repression of a restricted number of genes. The hormone response elements and associated factor elements are called hormone response units³ (Fig. 1).

Group II hormones are water soluble and initiate a response by binding to a receptor located in the plasma membrane. Cyclic AMP (3', 5'-adenylic acid) a ubiquitous nucleotide derived from adenosine triphosphate (ATP) through the action of the enzyme adenylate cyclase plays a vital role in the action of several hormones. The intracellular level of cAMP is increased by hormones of this group and they do so through unique receptors converging on a single class of adenylate cyclase molecules.

Endocrinal disruptors

There is increasing evidence that some industrial chemicals including solvents may interfere with complex and carefully regulated hormonal messenger system of our bodies. The public, scientific and regulatory concern regarding the potential adverse health impacts of exposure to endocrine-disrupting chemicals have been reviewed by several reports published in early decades⁴⁻¹⁶. However, all these reports were limited to the single system or the particular endocrinal disrupting xenobiotic. During 31 March to 1 April 2004, a workshop was conducted in Mallorca, Spain on multi-organic risk assessment of EDCs. The objective of workshop was to develop risk assessment methods considering mixed exposures and low dose effects¹⁷.

The possible disruption of endocrine systems by environmental chemicals and the effects on human health have become major topics of discussion and



Fig. 1—Mechanism of action of lipophilic hormones (ref. 3)

research in past few years¹⁸. The issue of "environmental endocrine disruptors" is now a key topic on the agenda of groups in government, industry and toxicologists. In order to establish a common basis for discussion and to enable the identification of active substances, it is important to develop a precise definition of an endocrine disruptor¹⁹. There have been several attempts to describe the phrase but there is as yet no universally accepted definition of endocrine disruptor. Kavlock²⁰ defined an endocrinal disruptor as "an exogenous agent that interferes with the production, release, transport, metabolism, binding, action, elimination of natural hormones in the body responsible for the maintenance of homeostasis and the regulation of developmental process"²¹. However, another definition recently accepted during an European workshop on endocrine disruptors states:

"An endocrine disruptor is an exogenous substance or mixture that causes adverse health effects in an intact organism or its progeny, subsequent to change in endocrine function"²².

A large number of man-made chemicals that have been released into the environment as well as a few natural ones, have the potential to disrupt the endocrine system of animals and humans. There are persistent, bioaccumulative organohalogen compounds that include some fungicides, herbicides, insecticides and industrial chemicals and other synthetic products. Wildlife including birds and fish, zebrafish, shellfish and mammals are also affected by these compounds^{7,23-27}. Due to their similarity during early embryonic development, researchers used the data acquired on these species to make predictions about endocrine disrupting effects of chemicals on humans.

Toxicity of organic solvents

Organic solvents refer to a group of volatile compounds or mixtures with low molecular weight that are relatively stable chemically and that exist in the liquid state at temperatures of approximately $0^{\circ} - 250^{\circ}$ C ($32^{\circ} - 482^{\circ}$ F). Common organic solvents are classified as aliphatic hydrocarbons, cyclic hydrocarbons, aromatic hydrocarbons, halogenated hydrocarbons, ketones, amines, esters, alcohols, aldehydes and ethers. Many common solvents often exist as mixtures or blends of chemical compounds²⁸. Organic solvents are used for extracting, dissolving or suspending materials such as fats, waxes and resins that are not soluble in water. The removal of the solvent from a solution permits the recovery of the solute intact with its original properties²⁹. Solvents are used in paints, adhesives, cosmetics, glues, polymers, plastics, textiles, printing inks, agricultural products and pharmaceuticals²⁸. Approximately 9.8 million workers are potentially exposed to organic solvents³⁰. They share properties such as lipophilicity and volatility. Some of solvents are hydrophilic or less volatile. In general, inhalation and skin absorption represent significant routes of exposure to organic solvents. The uptake of vapour by inhalation is a simple physical process, the molecules diffuse from the alveolar space into the blood, where they dissolve. The molecules partition between two media-between air and blood during the absorptive phase and between blood and other tissues during the distribution phase. The more soluble a vapour is in the blood, the more of it will be absorbed into the blood during each respiratory cycle. The amount of vapour taken up by various tissues depends on the affinity of the organic solvent for each tissue. Thus the rate of the absorption of organic solvents in the lungs and distribution of various tissues including endocrine glands are variable and depend on the blood-gas and fat-blood partition coefficients³¹. Major organic solvents proved or suspected as endocrinal disruptors are summarized in Table 1.

Specific endocrine system as targets of organic solvents

Endocrine system can be affected by various organic compounds. Some endocrine organs appear to be more sensitive to toxic agents including organic solvents and often lead to multiple disruptions in the hormonal balance of the organism.

Generally chemically induced changes that affect the endocrinal organs seldom manifest after a single exposure to a toxic agent. Rather, solvents that have the potential to exert deleterious effects upon the endocrine system ordinarily require longer durations of exposure and repeated administrations. Depending upon the particular chemical, the sites of action may differ in their sensitivity to toxic agents. A physiological hierarchy of control regulates and counter-regulates the homeostasis of the endocrine system through a series of feed back mechanisms. Feed-back is the modulation of output by an endproduct or product (hormone or metabolite). A negative feed back mechanism inhibits an endocrine

1.	Benzene	22.	n-hexane
2.	Benzyl chloride	23.	Thiols
3.	Ethyl acetate	24.	1,1,1-trichloroethane
4.	Carbon disulfide	25.	1,1,2-trichloroethane
5.	Carbon tetra- chloride	26.	Trichloroethylene
6.	Chloroethane	27.	Pentachloroethane
7.	Chloroform	28.	Styrene
8.	Chloroprene	29.	1,1,1,2-tetrachloroethane
9.	Cresol	30.	1,1,2,2-tetrachloroethane
10.	Di-2-ethyl hexyl phthalate	31.	Vinyl acetate
11.	Dioxin	32.	Xylene
12.	Epichlorohydrin (Suspected)	33.	Phenols
13.	Ethylene dibro- mide	34.	Ethyl benzene
14.	Ethylene dichlo- ride	35.	1,4-bio[2-(3,5- dichloropyridyloxy)]benzene
15.	Furfuryl alcohol	36.	Polychlorinated biphenyls (PCBs)
16.	Glycol ethers	37.	m-dinitrobenzene
17.	Isopropyl alco- hol	38.	Bromo trichloroethane
18.	Ketones	39.	Carbon tetra bromide
19.	Methyl alcohol	40.	Benzyl bromide
20.	Nitriles	41.	Benzene hexachloride
21.	Toluene	42.	Tobacco smoke (containing many organic solvents)

Table 1-List of some organic solvents as endocrinal disruptors

pathway whereas positive feed back loop enhances an endocrine response. Feed back mechanisms can further be modulated by other endocrine systems that are either pulsatile, cyclical or stimulated through other mechanisms².

The concept of the endocrine system is that endocrine cells release a hormone that is transported to a receptor site in a target tissue where the hormone binds and exerts its biological effects. Perturbation of a target organ by a chemical can be mediated through central nervous system by interfering with trophic hormone secretion and also occur by a chemical acting directly upon the particular target organ. Further, hepatic microsomal enzyme system responsible for endogenous steroid metabolism can be affected by organic solvents^{32,33}.

Adrenal toxicity

Adrenal cortex is a multifunctional steroidogenic organ. Adrenal glands have many features which render them susceptible to toxicological insult including high membrane content of unsaturated fatty acid, high rate of blood flow, free radical generation during steroid biosynthesis, potential for bioactivation of xenobiotics by cytochrome P₄₅₀ enzymes, mechanism for uptake and storage of lipoprotein. The adrenal cortex secretes a range of steroid hormones, which are produced from cholesterol by reactions mainly catalyzed by cytochrome P_{450}^{34} . The enzymes of steroid biosynthesis are targets for various toxicants including or-ganic solvents^{35,36}. Adrenal insufficiency as a result of organic solvent exposure causes severe changes in electrolytes and carbohydrate metabolism, leading to circulatory collapse, hypoglycaemia, coma and sometime death. Trichloroethylene toxicity of adrenal glands has been described by Mazza and Brancaccio³⁷. The cortex and medulla integrate many biochemical changes in the internal and external environment. The adrenal cortex is vulnerable to chemical lesions³⁸. Various solvents cause severe damage to adrenal cortex. Carbon tetrachloride alters the adrenal gland secretion and causes adreno-cortical necrosis³⁹. Acrylonitrile produces adrenal necrosis in animals⁴⁰. Chemically induced dysfunction of adrenal gland has been reported⁴¹. Toluene elevates corticosterone on inhalation⁴². The effects of several drugs and chemicals on the structure of adrenal gland were described by Ribelin⁴³. Benzene and toluene exposure stimulated hypothalamic pituitary adrenocortical activity. Elevated corticosterone has also been reported to inhibit IL-2 production and impair immunocompetence⁴⁴. Transplacental toxicity of 3-methylsulphonyl-DDE in the developing adrenal cortex in mice was reported by Jonsson⁴⁵. Cytochrome P_{450} catalyzed the binding of 3-methyl-sulfonyl-DDE and o,p-DDD in human adrenal zona fasciculata and zona reticularis⁴⁶. Toluene inhalation induces the adrenocortical hypertrophy in rat⁴⁷. An adrenal dependent leucopenia after short term exposure to air borne irritants viz. acetic acid, benzyl chloride, 1,1-dichlorobenzene, ethyl acetate, ethyl acrylate, formaldehyde, isophorane, mesityloxide, phenol, styrene, toluene and vinyl toluene was reported by Brondeau⁴⁸. Adrenal cortex and medulla function may be altered by benzene, dioxane, tetrachloroethylene, PCB and TCDD also⁴⁹. Recently, the effects of endocrine disrupting chemicals on adrenal function have been studied⁵⁰.

Thyroid toxicity

The function of the thyroid is to elaborate, store and release thyroid hormones into the bloodstream. Available literatures on the toxic effects of various organic solvents shows that poly halogenated biphenyls affect thyroid gland metabolism in a number of animals⁵¹. Hyperthyroidism is known to augment hepatic sensitivity to chloroform⁵². The effect of carbon tetrachloride is aggravated by the administration of thyroxine⁵³. Specific chemicals appear to have a direct effect on the thyroid gland resulting in genetic damage that leads to cell transformation and tumor formation. Examples of thyroid initiaters include 2acetylamino fluorine, dichlorobenzidine, N-methyl-Nnitrosourea, methyl cholanthrene and polycyclic hydrocarbons. Multiple sites of hypothalamic pituitary thyroid triad exhibited by chemicals that develop an increased incidence of hyperplastic and/or neoplastic lesions are shown in Fig. 2.

Study on the inhalation toxicity of methanol, toluene and mixtures of both the chemicals revealed that thyroid gland in females appeared to be a target organ, although the changes were confined to a mild and occasionally moderate reduction in follicular size⁵⁵. Thyroid gland follicular cell hyperplasia was also observed after the exposure to ethyl benzene⁵⁶.

1,4-bio[2-(3,5-dichloropyridyloxy)] benzene promotes the thyroid carcinogenesis in rat⁵⁷. Mixtures

of 3,3',4,4',5-pentachlorobiphenyl and 2,2'4,4',5,5'hexachlorobiphenyl in female Harlen Sprague-Dawley rats alters the serum thyroid hormone levels⁵⁸. 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) acts as a potent and persistent thyroxine agonist^{59,60}. Schmutzler⁶¹ reported the effect of endocrine active compounds on thyroid hormone levels in serum and its action in liver, heart and kidney. Thyroid toxicity due to subchronic exposure to a complex mixture of sixteen organo chlorines has also been documented⁶².

Parathyroid toxicity

Parathyroid glands are composed of a single cell concerned with the biosynthesis type of parathormone. Relatively few solvents are known to parathyroid toxicity. Di-propylene glycol cause causes the secondary lesions in the parathyroid of rats⁶³. Our studies have shown that male parathyroidectomy modulates liver injury caused by CCl₄ in rat⁶⁴⁻⁶⁸. Solvent toxicity in parathyroid glands, however, remains unknown.

Pancreas toxicity

The endocrine pancreas by secreting the polypeptide hormones i.e. insulin and glucagon controls blood



Fig. 2—Multiple sites of hypothalamic-pituitary-thyroid triad exhibited by chemicals that develop an increased incidence of hyperplastic and/or neoplastic lesions (ref. 54).

glucose levels. These hormones have opposite actions; while insulin promoting glucose uptake and utilization by tissues, the glucagon facilitating glycogenolysis and gluconeogenesis. Abnormal regulation of blood glucose leading to hyperglycemia is a characteristic of diabetes, a multifaceted disease characterized by deficiencies in the function of the endocrine pancreas particularly the insulin secreting β -cells⁶⁹. Heavy alcohol consumption is a potential risk factor for induction of pancreatitis^{70,71}. In an earlier study, functional state of pancreas in workers engaged in industries using benzene was described⁷². Increased bile duct pancreatic fluid flow in benzene and halogenated benzene treated rats was also studied⁷³. The occupational exposure to polycyclic aromatic hydrocarbons and k-ras activation in human exocrine pancreatic cancer was studied⁷⁴. A study on cancer incidence of pancreas among Finnish workers exposed to aromatic hydrocarbons (styrene, toluene, xylene) has been made⁷⁵. The effects of benzoic acid and its analogue on insulin and glucagon secretion were studied in sheep⁷⁶. Gasoline containing benzene, toluene 1,3butadiene, ethyl benzene, xylene, trimethyl pentane, methyltertbutylethane (MTBE) produced the cancer of pancreas in workers engaged in oil refinary and petrochemical industry^{77,78}. A fatal case of oral ingestion of toluene and its distribution in tissues including pancreas was also studied⁷⁹. In vitro effects of polyphenol on activity expression and secretion of pancreatic bile salt dependent lipase were described by Sbarra and coworkers⁸⁰. A very interesting study, described the chronic pancreatitis in cigarette smoking patients⁸¹. The study revealed expression of p-53 protein in chronic pancreatitic patients. 2,4-dimethoxy-2-methylnaphthalene (DMN) potentiated the apoptosis of pancreatic acinar cells after induction of menadione⁸². Endocrine pancreas deserves special attention since exposure to many xenobiotics can contribute in the etiology of diabetes mellitus.

Pituitary gland toxicity

Pituitary functions can be altered by many endogenous and exogenous agents⁸³. Generally chemical induced changes that affect pituitary target organ relationships seldom manifest after a single exposure of a toxic agent. Polychlorinated biphenyls (PCBs) are persistent food contaminants that have adverse effects on the pituitary gland⁸⁴. Another study described the effect of chloroform narcosis and other narcosis on the activity of basic carboxy-peptidases in the hypothalamic- pituitary- adrenal axis in rats⁸⁵.

However, bromodichloromethane decreased the incidence of pituitary gland tumors in female rats and in female mice⁸⁶. Ethanol induces oxidative damage in the pituitary gland⁸⁷. Chronic inhalation of hexane by female F-344 rats and B6C3F1 mice produced pituitary adenomas⁸⁸. Role of pituitary hormones in the regulation of hepatic cytochrome $P_{450}2E1$ in rats and mice was also studied⁸⁹. The fetal pituitary gonadotropin was reported to be an initial target in the impairment of cholesterol transportation and steroidogenesis in rats⁹⁰. Cell death in ALT-20 pituitary cells was induced by 2,3,7,8-TCDD⁹¹. Induction of 7-ethoxyresorufin-o-deethylase activity by chlorinated hydrocarbons and polycyclic aromatic hydrocarbons in cell lines was reported from the rainbow trout pituitary⁹². Effects of various occupational agents' viz. benzene, dioxane, styrene, tetrachloroethylene and toluene on the endocrine glands including pituitary were also reported⁴⁹. TCDD affects the pituitary gland by various ways, however, information on toxicity of other industrial solvents is wanting⁹³⁻¹⁰⁰.

Testis toxicity

In 1775, an English physician, Percival Pott reported a high incidence of scrotal cancer among chimney sweep workers¹⁰¹. One of the key questions most frequently asked from toxicologists investigating testicular toxicity is "does this mean that reproductive function will be affected"¹⁰². There are several organic solvents which manifest testicular damage in experimental animals. man and DNB (mdinitrobenzene) is notorious for causing the testicular damage. The metabolism of DNB in testicular cells (by Sertoli cells) is shown in Fig. 3.

Within the testes, members of the cytochrome P_{450} play an important role in solvent metabolism¹⁰³. Chronic exposure to hexane or methyl butyl ketone results in testicular damage¹⁰⁴. 7,12-dimethyl benz(a)anthracene (DMBA) is a polycyclic aromatic hydrocarbon that causes damage to the testis ¹⁰⁵. 2,5hexanedione and toxicity of aliphatic hydrocarbon has workers^{104,106-111} several been studied bv Spermatozoon is target for reproductive toxicants¹¹². The nucleus of spermatozoa may be targeted by reactive chemicals that damage DNA. Reactive oxygen species may alter the membrane function and damage DNA in the nucleus of spermatozoa (Fig. 4).



Fig. 3—Metabolism of m-dinitrobenzene by Sertoli cells (ref. 102)



Fig. 4—DNA damage by ROS in the spermatozoon (ref. 112).

Further studies on the testicular toxicity caused by industrial solvents will be important for problems related with human reproduction.

Benzene, toluene and xylene affect the semen quality and function of accessory gland of workers exposed to these chemicals¹¹³. Co-administration of toluene and xylene antagonized the testicular toxicity¹¹⁴. The effect of thinner and its main components, toluene, xylene, methanol and ethyl acetate on testes in male rats was described¹¹⁵. nhexane causes the testicular atrophy in rats¹¹⁶. Halogenated compounds (carbon tetrachloride. bromotrichloroethane, carbon tetrabromide, p-bromo benzyl bromide, benzyl bromide) are the potential inducers of lipid peroxidation in rat testis¹¹⁷. A known carcinogen ethylbenzene induces the testicular tumor¹¹⁸. Benzene induces histopathological lesions in testes¹¹⁹. An earlier review described the toxicity of benzene, glycol ethers and carbontetrachloride¹²⁰. hexachloride Benzene also causes the

histopathological and biochemical changes in testes¹²¹. The action of paradicyanobenzene on the testes of mouse was studied¹²². Hexafluoroacetone affect the Leydig cell steroidogenesis and spermatogenesis in rat¹²³. Occupational exposure and sniffing of toluene based organic solvents inhibit testosterone synthesis¹²⁴. Toluene also induces testes¹²⁵. the oxidative DNA damage in Trichloroethylene after inhalation causes testicular toxicity in rats^{126,127}. Higher concentration of benzene could cause damage to the sperm DNA in industrial workers¹²⁸

Ovary toxicity

The ovaries are target organs for injury caused by many chemicals¹²⁹. The female reproductive system is complex and multifactorial. The ovary is the central component of the hypothalamic pituitary-ovarian axis. It functions cyclically to produce a single oocyte. The follicle is the basic functional unit of the ovary and consists of an immature oocyte surrounded by multiple layers of specialized follicular cells, the theca cells and the granulosa cells¹³⁰. There are various organic solvents which cause damage to the ovaries and its functions. 1-Bromopropane induces ovarian dysfunction in non-pregnant female rats associated with disruption in follicular growth process 131 . Benzene and its analogues affect the luteal function of female workers in petrochemical industries¹³². Ovarian toxicity and carcinogenicity of several

organic compounds viz. 1,3-butadiene, 4vinylcyclohexane, vinylcyclohexane diepoxide, nitrofurantoin, nitrofurazone and benzene has also been described by Maronpot¹³³ who reported typical non neoplastic ovarian changes i.e. hyperplasia, atrophy, follicular necrosis and tubular hyperplasia and granulose cell tumors and benign mixed tumors. Benzene fumes also affect the functional activity of ovaries of rats¹³⁴. Dioxin inhibits follicular development¹³⁵. Smoking induces oxidative stress inside the graffian follicle¹³⁶. The effect of chronic inhalation of toluene and dioxane on activity of free radical processes in rat ovaries were also studied¹³⁷. Mixtures of chlorinated alkanes and alkanes consisting of chloroform, 1,1-dichloroethane, 1,1dichloroethylene, 1,1,1-trichloroethane, trichloroethylene and tetrachloroethylene increased the ovary total weights in female ICR mice¹³⁸. Women smokers and women exposed to environmental tobacco smoke containing various organic vapours have reduced ovarian function as evidenced by an earlier menopause, reduced follicular numbers, decreased levels of circulating estradiol and decreased conception rates¹³⁹.

Conclusion

Endocrine and hormonally modulated toxicity of xenobiotics have been paid less attention in comparison to other areas of toxicology research. List of chemicals causing primary, secondary and indirect endocrine toxicity hardly mention industrial solvents. Each endocrine gland deserves much more and detailed attention given hitherto. Diversity and complexity of pituitary gland and its interrelationship with brain provide many opportunities for xenobiotics to produce toxicity. Evidence for direct effects of solvents on thyroid and parathyroid glands are also lacking. Pathology of adrenal cortex has been studied but toxicological effects on medulla have rarely been investigated. No major studies have been made to estimate the role of environmental poisons in male reproductive toxicity. Close association between occupational exposure to industrial solvents and infertility necessitates to develop a reliable animal models of testicular dysfunction. Developmental effects of solvents further cause considerable public health concern. Response of endocrine pancreas to toxic injury in terms of apoptosis, necrosis, carcinogenesis, neuroendocrine tumor syndromes, insulinoma, glucagonoma, and multiple endocrine

neoplasia syndromes is known for xenobiotics other than organic solvents. In addition, the effects of industrial solvents on cytokines need special attention. To summarize, it is not only the endocrine disruption but endocrine toxicity of industrial chemicals as a whole that needs to be revisited in terms of modern knowledge on hormones and their disorders.

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References

- 1 Hadley ME, The vertebrate endocrine system, in *Endocrinology*, 4th Edition (Prentice Hall, NJ) 1996, 14.
- 2 Thomas JA & Thomas MJ, in *Endocrine toxicology* edited by B Ballantyne, T Marrs and T Syversen, *General and applied toxicology* (Macmillan Reference Ltd., London) 2 (1999) 979.
- 3 Granner DK, *Hormonal action, principles and practice of endocrinology and metabolism*, edited by K L Becker, III ed. (Williams and Wilkins, Lippincott, Baltimore, MD, USA) 2001, 42.
- 4 Henderson BE, Benton B, Cosgrove M, Baptista J, Aldrich J, Townsend D, Hart W & Mack TM, Urogenital tract abnormalities in sons of women treated with diethylstilbestrol, *Pediatrics*, 58 (1976) 505.
- 5 Bibbo M, Gill WB, Azizi F, Blough R, Fang VS, Rosenfield RL, Schumacher GF, Sleeper K, Sonek MG & Wied GL, Follow-up study of male and female offspring of DES-exposed mothers, *Obstet Gynecol*, 49 (1977) 1.
- 6 Carlsen E, Giwercman A, Keiding N, Skakkabaek NE, Evidence for the decreasing quality of semen during the past fifty years, *Br Med J*, 305 (1992) 609.
- 7 Colborn T, Vom Saal FS & Soto AM, Developmental effects of endocrine-disrupting chemicals in wildlife and humans, *Environ Health Perspect*, 101 (1993) 378.
- 8 Sharpe RM & Skakkebaek NE, Are estrogens involved in falling sperm counts and disorders of the male reproductive tract, *Lancet*, 341 (1993) 1392.
- 9 Davis DL, Bradlow HL, Wolff M, Woodruff T, Hoel DG & Anton-Culver H, Medical hypothesis: Xenoestrogens as preventable causes of breast cancer, *Environ Hlth Perspect*, 101 (1993) 372.
- 10 Giusti RM, Iwamoto K & Hatch EE, Diethylstilbestrol revisited: A review of the long-term health effect, *Ann Intern Med*, 122 (1995) 778.
- 11 Newbold R, Cellular and molecular effects of developmental exposure to diethylstil bestrol: implications for other environmental estrogens, *Environ Health Perspect*, 103 (1995) 83.
- 12 Davis DL & Bradlow HL, Can environmental estrogens cause breast cancer, *Sci Am*, 273 (1995) 166.
- 13 Safe S, Dietary and environmental estrogens and antiestrogens and their possible role in human disease, *Environ Sci Pollut Res*, 1 (1994) 29.
- 14 Safe S, Environmental and dietary estrogens and human

health- Is there a problem? *Environ Hlth Perspect*, 103 (1995) 346.

- 15 Safe S, Endocrine disruptors and human health: Is there a problem, *Toxicology*, 205 (2004) 3.
- 16 Ahlborg UG, Lipworth L, Titusernstoff L, Hsieh CC, Hanberg A, Baron J, Trichopoulos D & Adami HO, Organochlorine compounds in relation to breast cancer endometrial cancer and endometriosis: an assessment of the biological and epidemiological evidence, *Crit Rev Toxicol*, 25 (1995) 463.
- 17 Wutke W, Multiorganic risk assessment-summary of a CREDO cluster workshop, *Toxicology*, 205 (2004) 1.
- 18 Harrison PT, Holmoes P & Humfrey CD, Reproductive health in humans and wild life: are adverse trends associated with environmental chemical exposure? *Sci Total Environ*, 205 (1997) 97.
- 19 Humfrey CDN & Smith LL, Endocrine disrupting chemicals: The evidence for human health effects, in *Endocrine and hormonal toxicology*, edited by P.W Harvey, K.C. Rush and A.Cockburn (John Wiley & Sons, NY, USA) 1999, 421.
- 20 Kavlock RJ, Research needs for risk assessment of health and environmental effects of endocrine disruptors: A review of the US-EPA-sponsored workshop, *Environ Health Perspect*, 104 (1996) 715.
- 21 Fisher JS, Are all EDC effects mediated via steroid hormone receptors? *Toxicology*, 205 (2004) 33.
- 22 IPCS, The international programme on chemical safety (IPCS): Global assessment of the state-of-the-science of endocrine disruptors, WHO/PCS/EDC/02, (2002) 2.
- 23 Colborn T & Clement C, Chemically induced alterations in sexual and functional development: The wildlife/human connection, Advances in modern environmental toxicology (Princeton Scientific Publishing Co. Inc., NY, USA) 1992, 12.
- 24 Wester PW, Vander Ven LTM, Van Den Brandhof ES & Vos JH, Identification of endocrine disruptive effects in aquatic environment: A partial life cycle study in Zebrafish. Report. 640920, (RIVM, the Netherlands) 2003, 112.
- 25 Wester PW, Vander Ven LTM & Vos JG, Comparative toxicological pathology in mammals and fish: Some examples with endocrine disuptors, *Toxicology*, 205 (2004) 27.
- 26 Vanden Belt K, Wester P, Vander Ven LTM, Verheyen R, Witter H, Time dependent effects of ethynylestradiol on the reproductive physiology in Zebrafish (Danio rerio), *Environ Toxicol Chem*, 21 (2002) 767.
- 27 Salazar V, Castillo C, Ariznavarreta C, Campon R & Tresguerres JAF, Effect of oral intake of dibutyl phthalate on reproductive parameters of long Evans rats and prepubertal development of their offspring, *Toxicology*, 205 (2004) 131.
- 28 WHO, Nordic Council of Ministers, Organic solvents and the central nervous system. EH5. (World Health Organization and Nordic Council of Ministers,

Copenhagen, Denmark) 1985, 1.

- 29 Considine DM, Van Nostrand's scientific encyclopedia, 5th edition (Van Nostrand Reinhold Company, New York) 1976, 2048.
- 30 NIOSH, National occupational hazard survey 1972-1974, U.S. Department of Health, Education, and Welfare, Public Health Service, Center for Disease Control, National Institute for occupational safety and health, DHEW (NIOSH, Cincinnati, OH) Publication, 1977, 78.
- 31 Sato A & Nakajima T, Pharmacokinetics of organic solvent vapours in relation to their toxicity, *Scand J Work Environ Health*, 13 (1987) 81.
- 32 Verma Y & Rana SVS, Effects of progesterone on benzene toxicity in rats, *Arh Hig Rada Toksikol*, 59 (2008) 1.
- 33 Verma Y & Rana SVS, Modulation of CYP₄₅₀2E1 and oxidative stress by testosterone in liver and kidney of benzene treated rats, *Indian J Exp Biol*, 46 (2008) 568.
- 34 Hinson JP & Raven PW, Adrenal toxicology, in *Endocrine* and hormonal toxicology, edited by P W Harvey, K C Rush and A Cockburn (John Wiley and Sons, NY, USA) 1999, 67.
- 35 Hinson JP & Raven PW, Adrenal morphology and hormone synthesis and regulation, in *The adrenal in toxicology*, edited by P W Harvey (Taylor and Francis, London) 1996, 23.
- 36 Allera A, Lo S, King I, Steglich F & Klingmuller D, Impact of androgenic/antiandrogenic compounds (AAC) on human sex steroid metabolizing key enzymes, *Toxicology*, 205 (2004) 75-85.
- 37 Mazza V & Brancaccio A, The adrenal gland in experimental trichloroethylene poisoning, *Folia Med.* (Napoli), 50 (1967) 378.
- 38 Colby HD, Huang Y, Jiang KQ & Voigt J M, Toxicology of the adrenal cortex: role of metabolic activation, in *Endocrine toxicology*, 2nd edn., edited by J A Thomas and H D Colby (LippinCott-Raven, Philadelphia, PA) 1996, 81.
- 39 Colby HD & Longhurst PA, Toxicology of the adrenal gland, in *Endocrine toxicology*, edited by C K Atterwill and J D Flack (Cambridge University Press, Cambridge) 1992, 243.
- 40 Szabo S & Sandor Z, Chemically induced lesions in the adrenal cortex, in *Endocrine toxicology*, 2nd edn., edited by J A Thomas and H D Colby (Taylor and Francis, New York) 1997, 115.
- 41 Colby HD, Adrenal gland toxicity: Chemically induced dysfunction, *J Am Coll Toxicol*, 7 (1988) 45.
- 42 Andersson K, Nilsen OG, Toftgard R, Eneroth P, Gustafasson JA, Battistini N, & Agnati L F, Increased amine turnover in several hypothalamic noradrenaline nerve terminal systems and changes in prolactin secretion in the male rat by exposure to various concentrations of toluene, *Neurotoxicology*, 4 (1983) 43.
- 43 Ribelin WE, The effects of drugs and chemicals upon the structure of the adrenal gland, *Fundamental Appl Toxicol*, 4 (1984) 105.
- 44 Hseish GC, Sharma RP & Parker RD, Hypothalamicpituitary adrenocortical axis activity immune function after oral exposure to benzene and toluene, *Immunopharmacology*, 21 (1991) 23.
- 45 Jonsson J, Rodriquez-Martine H & Brandt I, Transplacental

toxicity of 3-methylsulphonyl-DDE in the developing adrenal cortex in mice, *Reprod Toxicol*, 9 (1995) 257.

- 46 Lindhe O, Skogseid B & Brandt I, Cytochrome P-450catalyzed binding of 3-methylsulfonyl-DDE and O,p'-DDD in human adrenal zona fasciculata/reticularis, *J Clin Endocrinol Metab*, 87 (2002) 1319.
- 47 Gotohda T, Tokunaga I & Kubo S, Toluene inhalation induced adrenocortical hypertrophy and endocrinological changes in rat, *Life Scie*, 76 (2005) 1929.
- 48 Brondeau MT, Bonnet P, Guenier JP, Simon P & de Ceaurriz J, Adrenal dependent leucopenia after short term exposure to various airborne irritants in rats, *J Appl Toxicol*, 10 (1990) 83.
- 49 Baccarelli A, Occupational agents and endocrine function: An update of the experimental and human evidence, *Med Lav*, 90 (1999) 650.
- 50 Hinson JP, Raven PW, Effects of endocrine-disrupting chemicals on adrenal function, *Best Pract. Res. Clin Endocrinol. Metab.* 20 (2006) 111.
- 51 Collins WT & Capen CC, Ultrastructural and functional alterations of the rat thyroid gland produced by polychlorinated biphenyls compared with iodide excess and deficiency and thyrotropin and thyroxine administration, *Virchows Arch B Cell Pathol*, 33 (1980) 213.
- 52 Paget GE, The effect of beryllium sulfate on chloroform induced hepatic necrosis in mice, *Toxicol Appl Pharm*, 3 (1961) 595.
- 53 Calvert DN & Brody TM, The effects of thyroid function upon carbon tetracholoride hepatotoxicity, *J Pharm Exp Ther*, 134 (1961) 304.
- 54 Capen CC, Toxic responses of the thyroid gland, edited by IG Sipes, CA Mcqueen, and AJ Gandolfi, *Comprehensive Toxicol* (Elsevier Ltd, North Holland) 10 (1997) 695.
- 55 Poon R, Chu I, Bjarnason S, Potvin M, Vincent R, Miller RB & Valli VE, Inhalation toxicity study of methanol, toluene and methanol/toluene mixtures in rats: effects of 28-days exposure, *Toxicol Ind Health*, 10 (1994) 231.
- 56 National Toxicology Programme, NTP toxicology and carcinogenesis studies of ethyl benzene (CAS No. 100-41-4) in F344/N rats and B6C3F1 mice (inhalation studies), *Natl Tox Prog Tech Rep Ser*, 466 (1999) 1.
- 57 Diwan BA, Henneman JR, Rice JM & Nims RW, Enhancement of thyroid and hepatocarcinogenesis by 1,4bis[2,-(3,5-dichloropyridyloxy)] benzene in rats at doses that cause maximal induction of CYP2B, *Carcinogenesis*, 17 (1996) 37.
- 58 National Toxicology Program, NTP toxicology and carcinogenesis studies of binary mixture of 3,3,4,4',5pentachlorobiphenyl (PCB 126) (CAS No. 57465-28-8) and 2,3'4,4'5-pentachlorobiphenyl (PCB118) (CAS No. 31508-00-6) in female Harlan Sprague-Dawley Rats (Gavage Studies), *Natl Tox Prog Tech Rep Ser*, 531 (2006) 1.
- 59 Mckinney JD, Fawkes J, Jordan S, Chae K, Oatley S, Coleman RE & Briner. 2,3,7,8-tetrachlorodibenzo-pdioxide (TCDD) as a patent and persistent thyroxine agonist: A mechanistic model for toxicity based on molecular reactivity, *Environ Health Persp*, 61 (1985) 41.

- 60 Schuur AG, Boekhorst FM, Brouwer A & Visser TJ, Extrathyroidal effects of 2,3,7,8-tetrachlorodibenzo-pdioxin on thyroid hormone turnover in male Sprague-Dawley Rats, *Endocrinology*, 138 (1997) 3727.
- 61 Schmutzler C, Hamann I, Hofmann PJ, Kovacs G, Stemmler L, Mentrup B, Schomburg L, Ambrugger P, Gruters A, Wuttke DS, Jarry H, Wuttke W & Kohrle J, Endocrine active compounds affect thyrotropin and thyroid hormone levels in serum as well as endpoints of thyroid hormone action in liver, heart and kidney, *Toxicology*, 205 (2004) 95.
- 62 Wade MG, Parent S, Finnson KW, Foster W, Younglai E, McMohan A, Cyr DG & Hughes C, Thyroid toxicity due to chronic exposure to a complex mixture of 16 organochlorins, lead and cadmium, *Toxicological Sci*, 67 (2002) 207.
- 63 Hooth MJ, Herbert RA, Hasemen JK, Orzech DP, Johnson JD & Bucher JR, Toxicology and Carcinogenesis studies of dipropylene glycol in rats and mice, *Toxicology*, 204 (2004) 123.
- 64 Rastogi S & Rana SVS, Influence of parathyroidectomy on liver glycogen in rat treated with carbontetrachloride, *Indian J Exp Biol*, 28 (1990) 794.
- 65 Rana SVS & Rastogi S, Influence of parathyroidectomy on fatty liver, *J Zool Soc India*, 41 (1990) 83.
- 66 Rana SVS & Rastogi S, Effect of carbontetrachloride on collagen in the liver of parathyroidectomized rat, *Zool Jb Anat*, 121 (1991) 259.
- 67 Rana SVS & Rastogi S, Effect of parathyroidectomy on calcium and phospholipase A2 in the liver of carbontetrachloride treated rats, *Physiol Chem Phys and Med NMR*, 23 (1991) 173.
- 68 Rana SVS & Rastogi S, Antioxidative enzymes in the liver of rat treated with carbontetrachloride after parathyroidectomy, *Physiol Chem Phys and Med NMR*, 25 (1993) 41.
- 69 Fischer LJ, Toxicity of the insulin secreting beta cell, edited by IG Sipes, CA Mcqueen, AJ Gandelfi, *Comprehensive Toxicol*, 10 (1997) 701.
- 70 Ding YX, Yang K, Chin WC, Ethanol augments elevated [Ca²⁺]C induced tryps in activation in pancreatic acinar zymogen granules, *Biochem Biophys Res Commun*, 350 (2006) 593.
- 71 Chrostek L, Cylwik B & Szmit Kowski M, Ethanol metabolism in pancreas and its role in alcoholic pancreatitis, *Pol Arch Med Wewn*, 114 (2005) 989.
- 72 Bezhenova RV, Functional state of the stomach, pancreas and liver in workers engaged in industries using benzene, *Gig Tr Prof Zabol*, 6 (1962) 35.
- 73 Yang KH, Peterson RE & Fujimoto JM, Increased bile duct pancreatic fluid flow in benzene and halogenated benzene treated rats, *Toxicol Appl Pharmacol*, 47 (1979) 505.
- 74 Alguacil J, Porta M, Kauppinen T, Malats N, Kogevinas M & Carrato A, Occupational exposure to dyes, metals, polyclic aromatic hydrocarbons and other agents and k-ras activation in human exocrine pancreatic cancer, *Int J Cancer*, 107 (2003) 635.
- 75 Anttila A, Pukkala E, Riala R, Sallmen, M & Hemminki K, Cancer incidence among finnish workers exposed to

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aromatic hydrocarbons, Int Arch Occup Environ Health, 71 (1998) 187.

- 76 Mineo H, Ohdate T, Fukumura K, Katayama T, Onaga T, Kato S & Yanaihara N, Effects of benzoic acid and its analogues on insulin and glucagon secretion in sheep, *Eur J Pharmacol*, 280 (1995) 149.
- 77 Mehlman MA, Dangerous properties of petroleum refining products: carcinogenicity of motor fuels (gasoline), *Terat Carcin Mutagen*, 10 (1991) 399.
- 78 Mehlman MA, Dangerous and cancer causing properties of products and chemicals in the oil refining and petrochemical industry. VIII. Health effects of motor fuels: Carcinogenicity of gasoline scientific update, *Environ. Res*, 59 (1992) 238.
- 79 Ameno K, Fuke C, Ameno S, Kiriu T, Sogo K & Ijiri I, A fatal case of oral ingestion of toluene, *Forensic Sci Int*, 41 (1989) 255.
- 80 Sbarra V, Ristorcelli E, Petit-Thevenin JL, Teissedre PL, Lombardo D & Verine A, *In vitro* polyphenol effects on activity, expression and secretion of pancreatic bile salt dependent lipase, *Biochem Biophys Acta*, 1736 (2005) 67.
- 81 Sliwinska-Mosson M, Milnerowicz H, Milnerowicz S & Rabczynski J, Immunohistochemical localization of p53 protein in smoking patients with chronic pancreatitis, *Przegl Lek*, 63 (2006) 941.
- 82 Cridle DN, Gillies S, Baumgartner-Wilson HK, Jaffar M, Chinje EC, Passmore S, Chvanov M, Barrow S, Gerasimenko OV, Tepikin AV, Sutton R & Peterson OH, Menadione induced reactive oxygen species generation via redox cycling promotes apoptosis of murine pancreatic acinar cells, *J Biol Chem*, 281 (2006) 40485.
- 83 Thomas MJ & Thomas JA, Hormone assays and endocrine function, in *Principles and methods of toxicology*, 3rd edn., edited by AW Hayes (Raven Press, New York) 1994, 1039.
- 84 Johansson C, Tofighi R, Tamm C, Goldoni M, Mutti A & Ceccatel S, Cell death mechanisms in AfT20 pituitary cells exposed to polychlorinated biphenyls (PCB126 and PCB153) and methyl mercury, *Toxicol Lett*, 167 (2006) 183.
- 85 Vernigora AN, Gengin MT, Mukhina ES & Mikhailova OE, Effect of chloroform and ether necrosis on the activity of basic carboxy peptidases in the hypothalamic-pituitaryadrenal axis in rats, *Ukr Biokhim Zh*, 74 (2002) 124.
- 86 National Toxicology Program, NTP Toxicology and carcinogenens studies of bromo di chloromethane (CAS No. 75-27-4) in F344/N rats and B6C3F1 mice (Gavage studies), *Natl Toxic Prog Tech Rep Ser*, 321 (1987) 1.
- 87 Ren JC, Zhu Q, Lapaglia N, Emanuele NV & Emanuela MA, Ethanol induced alterations in Rab proteins: possible implications for pituitary dysfunction, *Alcohol*, 35 (2005) 103.
- 88 Daughtrey W, Newton P, Rhoden R, Kirwin C, Haddock L, Duffy J, Keenan T, Richter W & Nicolich M, Chronic inhalation carcinogenicity study of commercial hexane solvent in F-344 and B6C3F1 mice, *Toxicol Sci*, 48 (1999) 21.
- 89 Hong JY, Ning SM, Ma BL, Lee MJ, Pan JM & Yang CS, Roles of pituitary hormones in the regulation of cytochrome P₄₅₀2E1 in rats and mice, *Arch Biochem Biophys*, 281 (1990) 132.

- 90 Mutoh J, Taketoh J, Okamura K, Kagawa T, Ishida T, Ishii Y & Yamada H, Fetal pituitary gonadotropin as an initial target of dioxin in its impairment of cholesterol transportation and steroidogenesis in rats, *Endocrinology*, 147 (2006) 927.
- 91 Huang P, Tofighi R, Emgard M & Ceccatelli S, Cell death induced by 2,3,7,8-tetrachlorodibenzo-p-dioxin (2,3,7,8-TCDD) in AfT-20 pituitary cells, *Toxicology*, 207 (2005) 391.
- 92 Tom DJ, Lee LE, Lew J & Bols NC, Induction of 7ethoxyresorufin-O-deethylase activity by planer chlorinated hydrocarbons and polycyclic aromatic hydrocarbons in cell lines from the rainbow trout pituitary, *Comp Biochem Physiol A Mol Integr Physiol*, 128 (2001) 185.
- 93 Pitt JA, Buckalew AR, House DE & Abbott BD, Adrenocorticotropin (ACTH) and corticosterone secretion by perifused pituitary and adrenal glands from rodents exposed to 2,3,7,8-tetra chlorodibenzo-p-dioxin (TCDD), *Toxicology*, 151 (2000) 25.
- 94 Bestervelt LL, Pitt JA, Nolan CJ, Cai Y, Piper DW, Dybowski JA, Dayharsh GA & Piper WN, *In vitro* 2,3,7,8tetra chlorobibenzo – p-dioxin interference with the anterior pituitary hormone adrenocorticotropin, *Toxicol Sci*, 44 (1998) 107.
- 95 Bestervelt LL, Pitt JA & Piper WN, Evidence for Ah receptor mediation of increased ACTH concentrations in primary cultures of rat anterior pituitary cells exposed to TCDD, *Toxicol Sci*, 46 (1998) 294.
- 96 Chaffin CL, Trewin AL, Watanabe G, Taya K & Hutz RJ, Alterations to the pitutary-gonadal axis in the peributertal female rat exposed in utero and through lactation to 2,3,7,8-tetrachlorodibenzo-p-dioxin, *Biol Reprod*, 56 (1997) 1498.
- 97 Li J, Wang L & Guo J, Primary nasal ectopic pituitary adenoma: a case report, *Chin Med J* 110 (1997) 731.
- 98 Bookstaff RC, Kamel F, Moore RW, Bjerke DL & Peterson RE, Altered regulation of pituitary gonadotropin releasing hormone (GnRH) receptor number and pituitary responsiveness to GnRH in 2,3,7,8-tetrachlorodibenzo-pdioxin treated male rats, *Toxicol Appl Pharmacol*, 105 (1990) 78.
- 99 Moore HP, Brion C, Chung KN, Lehmicke L, Rivas R & Quinn D, Protein secretion by constitutive and regulated pathways, *Soc Gen Physiol* Ser, 44 (1989) 189.
- 100 Gorski JR, Muzi G, Weber LW, Pereira DW, Arceo RJ, Iatropoulos MJ & Rozman K, Some endocrine and morphological aspects of the acute toxicity of 2,3,7,8tetrachlorodibenzo-p-dioxin (TCDD), *Toxicol Pathol*, 16 (1988) 313.
- 101 Schrader SM & Kesner JS, Male reproductive toxicology, in Occupational and environmental reproductive hazards edited by M Paul (Williams & Wilkins, USA) 1993, 3.
- 102 Foster PMD, Evaluation of a toxicant, edited by IG Sipes, CA Mcqueen, AJ Gandolfi, *Comprehensive Toxicol*, 10 (1997) 63.
- 103 Juchau MR, Substrate specificities and functions of the P450 cytochromes, *Life Sci*, 47 (1990) 2385.
- 104 Krasavage WJ, O'Donoghue JL, Divincenzo GD et al., The

relative neurotoxicity of methyl n-butyl ketone, n-hexane and their metabolites, *Toxicol Appl Pharmacol*, 52 (1980) 433.

- 105 Ford E & Huggins C, Selective destruction in testes induced by 7,12-dimethyl benz(a)anthracene, *J Exp Med*, 118 (1963) 27.
- 106 Holloway AJ, Moore HD & Foster PM, The use of *in vitro* fertilization to detect reductions in the fertility of male rats exposed to 1,3-dinitrobenzene, *Fundam Appl Toxicol*, 14 (1990) 113.
- 107 Spencer PS, Schaumburg HH, Sabri MI & Veronesi B, The enlarging view of hexacarbon neurotoxicity, *Crit Rev Toxicol*, 7 (1980) 279.
- 108 Couri D & Milks M, Toxicity and metabolism of the neurotoxic hexacarbons n-hexane, 2-hexanone, and 2,5hexanedione, Annu Rev Pharmacol, Toxicol, 22 (1982) 145.
- 109 Chapin RE, Morgan KT & Bus JS, The morphogenesis of testicular degeneration induced in rats by orally administered 2,5-hexanedione, *Exp Mol Pathol*, 38 (1983) 149.
- 110 Boekelheide K & Eveleth J, The rate of 2,5-hexanedione intoxication, not total dose, determines the extent of testicular injury and altered microtubule assembly in the rat, *Toxicol Appl Pharmacol*, 94 (1988) 76.
- 111 Boekelheide K, 2,5-hexanedione alters microtubule assembly. I. Testicular atrophy, not nervous system toxicity, correlates with enhanced tubulin polymerization, *Toxicol Appl Pharmacol*, 88 (1987) 370.
- 112 Perreault SD, *The mature spermatozoon as a target for reproductive toxicants*, edited by I G Sipes, CA Mcqueen, A J Gandolfi, *Comprehensive Toxicol* (Elsevier Ltd., North Holland) 1997, 165.
- 113 Xiao G, Pan C, Cai Y, Lin H & Fu Z, Effect of benzene, toluene, xylene on the semen quality and the function of accessory gonad of exposed workers, *Industrial Health*, 39 (2001) 206.
- 114 Yu IJ, Lee JY, Chung YH, Kim KJ, Han JH, Cha GY, Chung WG, Cha YN, Park JD, Lee YM & Moon YH, Coadministration of toluene and xylene antagonized the testicular toxicity but not the haematopoietic toxicity caused by ethylene glycol monoethyl ether in sprague. Dawley rats, *Toxicol Lett*, 109 (1999) 11.
- 115 Yamada K, Influence of lacquer thinner and some organic solvents on reproductive and accessory reproductive organs in the male rat, *Biol Pharm Bull*, 16 (1993) 425.
- 116 Nylen P, Ebendal T, Eriksdotter-Nilsson M, Hansson T, Henschen A, Johnson AC, Kronevi T, Kvist U, Sjostrand NO, Hogland G & Olson L, Testicular atrophy and loss of nerve growth factor immuno-reactive germ cell line in rats exposed to n-hexane and a protective effect of simultaneous exposure to toluene or xylene, *Arch Toxicol*, 63 (1989) 296.
- 117 Fraga CG, Leibovitz BE & Tappel AL, Halogenated compounds as inducers of lipid peroxidation in tissue slices, *Free Radic Biol Med*, 3 (1987) 119.
- 118 Chan PC, Hasemani JK, Mahlari J & Aranyi C, Tumor induction in F344/N rats and B6C3F1 mice following inhalation exposure to ethyl benzene, *Toxicol Lett*, 99 (1998) 23.

- 119 Ward CO, Kuna RA, Snyder NK, Alsaker RD, Coate WB, Craig PH, Subchronic inhalation toxicity of benzene in rats and mice, *Am J Ind Med*, 7 (1985) 457.
- 120 Kalf GF, Post GB & Snyder R, Solvent toxicology: recent advances in the toxicology of benzene, the glycol ethers, and carbon tetrachloride, *Annu Rev Pharm Toxicol*, 27 (1987) 399.
- 121 Dikshith TS, Datta KK, Kushwah HS & Raizada RB, Histopathological and biochemical changes in guinea pigs after repeated dermal exposure to benzene hexachloride, *Toxicology*, 10 (1978) 55.
- 122 Julian M, Fabre MT, Voisin MC, Rakotondrainibe A, Rumeau JL & Bouissou H, Action of para dicyano-benzene on the testes of mouse (pathogenic deductions concerning human granulomatous orchitis), *Pathol Biol*, 18 (1970) 725.
- 123 Gillies PJ & Lee KP, Effects of hexafluoroacetone on Leydig cell steroidogenesis and spermatogenesis in the rat, *Exp Mol Pathol*, 42 (1985) 353.
- 124 Yilmaz B, Canpolat S, Sandal S, Akpolat N, Kutlu S, Ilhan N & Kelestimur H, Paint thinner exposure inhibits testosterone synthesis and secretion in a reversible manner in the rat, *Reprod Toxicol*, 22 (2006) 791.
- 125 Nakai N, Murata M, Nagahama M, Hirase T, Tanaka M, Fujikawa T, Nakao N, Nakashima K & Kawanishi S, Oxidative DNA damage induced by toluene is involved in its male reproductive toxicity, *Free Radical Res*, 37 (2003) 69.
- 126 Kumar P, Prasad AK & Dutta KK, Steroidogenic alterations in testes and sera of rats exposed to trichloroethylene (TCE) by inhalation, *Hum Exp Toxicol*, 19 (2000) 117.
- 127 Kumar P, Prasad AK, Mani U, Maji BK & Dutta KK, Trichloroethylene induced testicular toxicity in rats exposed by inhalation, *Hum Exp Toxicol*, 20 (2001) 585.
- 128 Song B, Cai ZH, Li X, Deng LX & Zheng LK, Effect of benzene on sperm DNA, *Zhonghua Nan Ke Xue*, 11 (2005) 53.
- 129 Mattison DR, Plowchalk DR, Meadows MJ, Al-Juburi AZ, Gardy J & Malek A, Reproductive toxicity: male and female reproductive systems as targets for chemical injury, *Med Clin N Am*, 74 (1990) 391.
- 130 Barbera AR, Differentiation and function of the female reproductive system, edited by IG Sipes, CA, Mcqueen, A.J. Gandolfi, Comprehensive Toxicol (Elsevier Ltd., North Holland), 1997, 255.
- 131 Yamada T, Ichihara G, Wang H, Yu X, Maeda K, Tsukamura H, Kamijima M, Nakajima T & Takeuchi Y, Exposure to 1-bromo-propane causes ovarian dysfunction in rats, *Toxicol Sci*, 71 (2003) 96.
- 132 Chen H, Song L, Wang X & Wang S, Effect of exposure to low concentration of benzene and its analogues on luteal function of female workers, *Wei Sheng Yan Jiu*, 29 (2000) 351.
- 133 Maronpot RR, Ovarian toxicity and carcinogenicity in eight recent national toxicology program studies, *Environ Health Perspect*, 73 (1987) 125.
- 134 Matysiak VG, The effect of benzene fumes on the functional activity of the hypophysis, adrenals and ovaries

of white rats under experimental conditions. *God Zb Med. Fak Skopje*, 14 (1968) 98.

- 135 Heiden TK, Carven MJ & Hutz RJ, Inhibition of follicular development, vitellogenesis and serum 17 beta-estradiol concentrations in zebrafish following chronic, sublethal dietary exposure to 2,3,7,8-tetra chlorodibenzo-p-dioxin, *Toxicol Sci*, 90 (2006) 490.
- 136 Paszkowski T, The influence of smoking on the estradiol level in the preovulatory follicular fluid, *Ginekol Pol*, 72 (2001) 989.
- 137 Burmistrov SO, Arutyunyan AV, Stepanov MG, Oparina TI & Prokopenko UM, Effect of chronic inhalation of

toluene and dioxane on activity of free radical processes in rat ovaries and brain, *Bull Exp Biol Med*, 132 (2001) 832.

- 138 Wang FI, Kuo ML, Shun CT, Ma YC, Wang JD & Ueng TH, Chronic toxicity of a mixture of chlorinated alkanes and alkenes in ICR mice, *J Toxicol Environ Health-A*, 65 (2002) 279.
- 139 Vidal JD, Vande Voort CA, Marcus CB, Lazarewicz NR & Conley AJ, *In vitro* exposure to environmental tobacco smoke induces CYP1B1 expression in human luteinized granulosa cells, *Reprod Toxicol*, 22 (2006) 731.